



# Tips and Tricks for Product Development

Unisart® Membranes in  
Lateral Flow Test Platform

Simplifying Progress

**SARTORIUS**

# Tips and Tricks for Product Development

## Unisart® Membranes in Lateral Flow Test Platform



Description	Action	Example
<b>Membrane storage and handling conditions</b>	<ul style="list-style-type: none"><li>▪ T 15-25° C   max 60% relative humidity</li><li>▪ Very dry conditions may alter the wettability and handling properties, can be overcome by conditioning the membrane at 40-60% RH for approx. 12 h</li><li>▪ Keep membrane originally packed in protected, well aerated area</li><li>▪ Keep membrane away from open fire, any sources of heat, light and chemical vapors</li><li>▪ After unpacking, please avoid any direct contact with the membrane, also with materials that have the potential to release chemicals or additives into the membrane (e.g., cardboard, plastic)</li></ul>	
<b>Unisart® Nitrocellulose Membranes</b>	<ul style="list-style-type: none"><li>▪ Choice of capillary flow rate</li><li>▪ Highly viscous samples and fast readout time: Unisart® CN 95, CN 110</li><li>▪ High precision and longer readout time: Unisart® CN 140U, CN 140B, CN 150, CN 180</li></ul>	<p>95... → ...180</p>
<b>Line printing conditions</b>	<ul style="list-style-type: none"><li>▪ Speed: 50-60 mm/sec</li><li>▪ Dispensing rate: 1.0-1.25 µL/cm</li></ul>	
<b>Printing buffer e.g., for IgG</b>	<ul style="list-style-type: none"><li>▪ 50 mm Phosphate, 3% methanol, pH 7.4</li><li>▪ 5 mm borate buffer, 150 mm NaCl, 1% sucrose, pH 8.0</li></ul>	
<b>Protein fixing conditions</b>	Fix proteins at 37-60° C for at least 5-15 min immediately after printing. Vary with nature of reagents, printing buffer composition and membrane	



Find out more

[www.sartorius.com/en/products/oem/oem-membranes-and-devices/diagnostic-membranes/unisart-lateral-flow](http://www.sartorius.com/en/products/oem/oem-membranes-and-devices/diagnostic-membranes/unisart-lateral-flow)



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<b>Uneven lines</b>		
<ul style="list-style-type: none"> <li>Dispensing equipment</li> </ul>	<ul style="list-style-type: none"> <li>Verify dispensing equipment</li> <li>Adjust speed, volume, humidity (50–60% RH), drying conditions</li> <li>Install device to reduce electrostatic charges</li> </ul>	
<i>Wang, et al; J Memb Sci. 2008</i>		
<ul style="list-style-type: none"> <li>Conjugate pad</li> </ul>	<ul style="list-style-type: none"> <li>Verify dispensing and free release of reagents on conjugate pad</li> <li>Add small amounts of surfactant to support homogeneous flow and reduce non-specific binding (e.g., 100 mM Tris-buffer, 0.5% BSA, 0.25% Tween, 5% saccharose, pH 8.0)</li> </ul>	
<ul style="list-style-type: none"> <li>Buffer</li> </ul>	<p>Optimizing capture antibody buffer for each reagent</p> <ul style="list-style-type: none"> <li>Reduce ionic strength: Low ionic strength helps binding to NC</li> <li>pH +/- 1 from isoelectric point to decrease protein stability in solution</li> <li>Up to 5% alcohol (MeOH or EtOH) to support wetting and protein binding</li> <li>Add minute amounts of surfactant: can support line morphology and prevent non-specific binding</li> </ul>	
<ul style="list-style-type: none"> <li>Slitting</li> </ul>	<p>Sharp edges are essential for homogeneous front flow and even lines</p> <ul style="list-style-type: none"> <li>Adjust blades for different backing thickness (50 vs. 100 μm)</li> </ul>	

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